

# Model-based reconstruction of whole organ growth dynamics reveals invariant patterns in leaf morphogenesis

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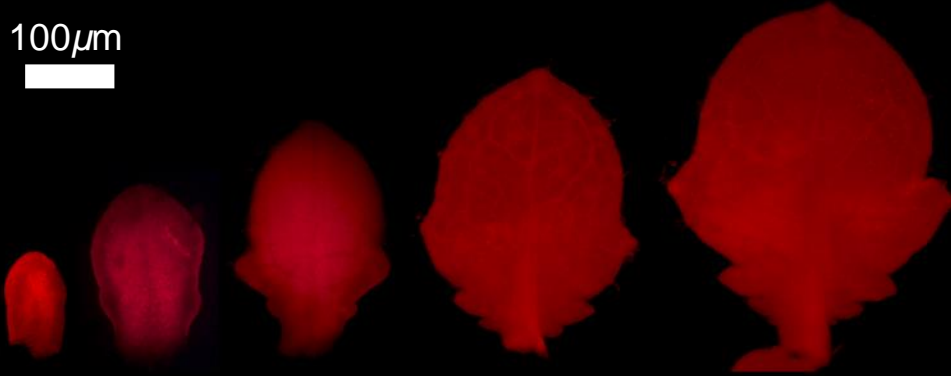
**INRAE**

AgroParisTech 

 **ijpb**

# Quantifying plant growth dynamics

100 $\mu$ m

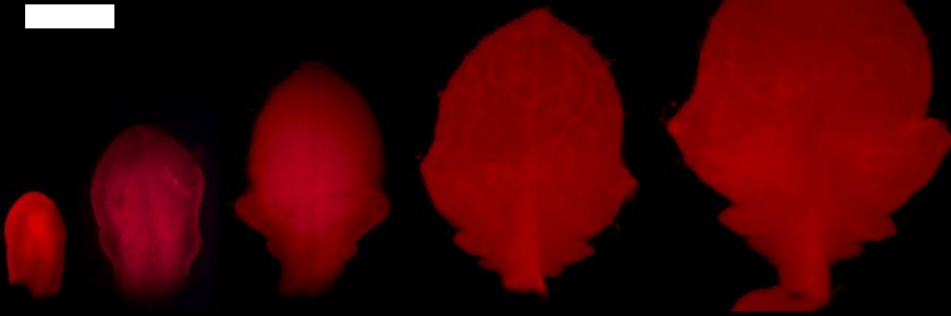


Organ morphogenesis implies long-term  
**spatio-temporal** processes

- **Global goal:** determine how/where/when these processes operate during growth
- **Intermediate goal:** quantify organ shape evolution during growth

# Quantifying plant growth dynamics

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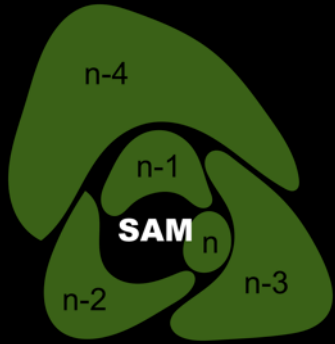


Organ morphogenesis implies long-term **spatio-temporal** processes

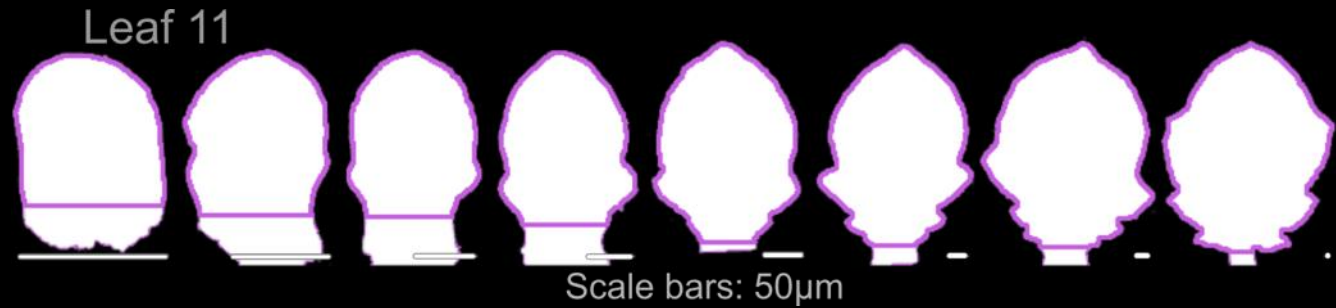
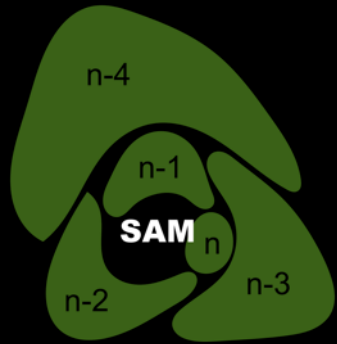
- **Global goal:** determine how/where/when these processes operate during growth
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- Some challenges:
  - organs are not always accessible/observable from early to late stages
  - organ size changes a lot (from micro- to macro-scales)
  - long-term observation may significantly impact growth itself
  - multiple observations must be done/integrated to face variability

# Quantifying plant growth dynamics: the leaf as a model

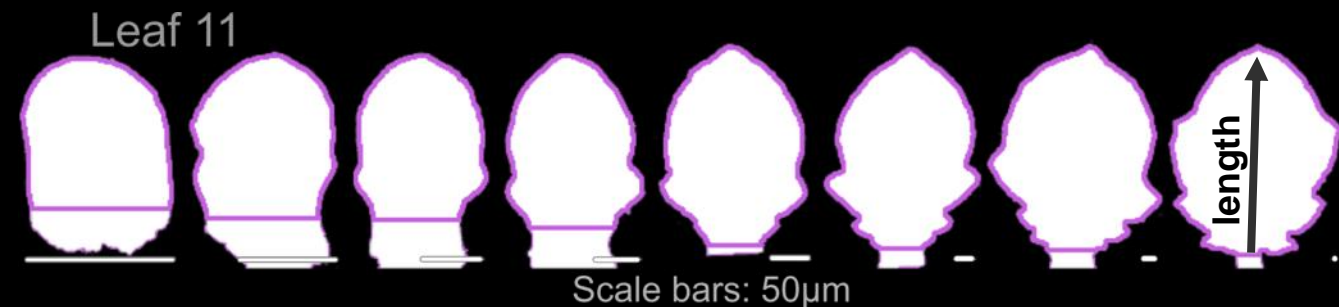


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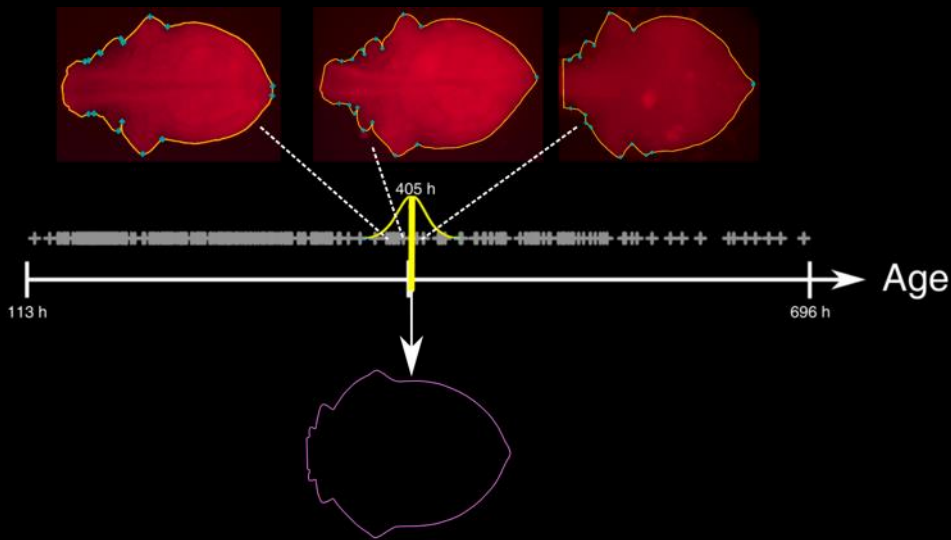


# Quantifying plant growth dynamics: *MorphoLeaf*

Biot et al. 2016

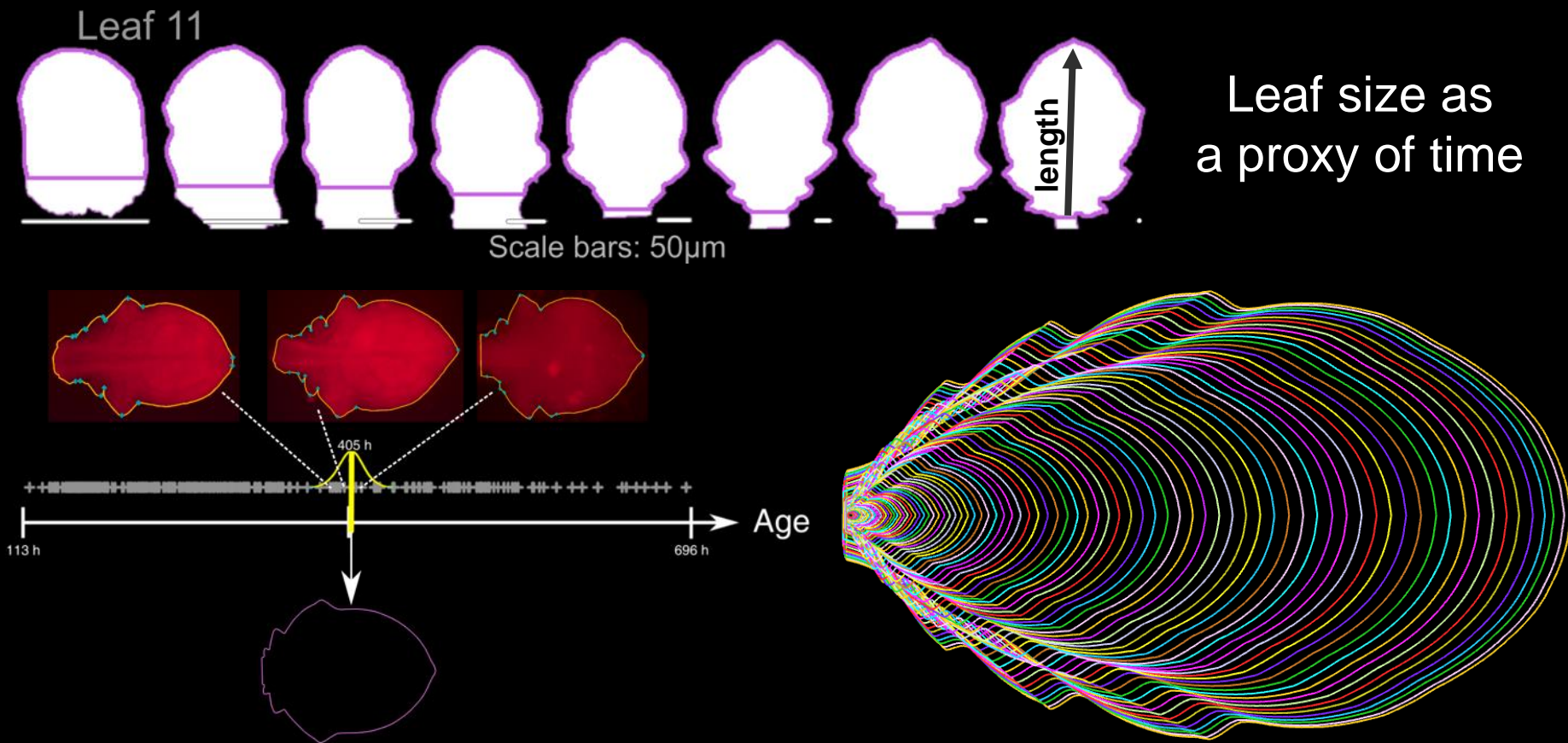


Leaf size as  
a proxy of time



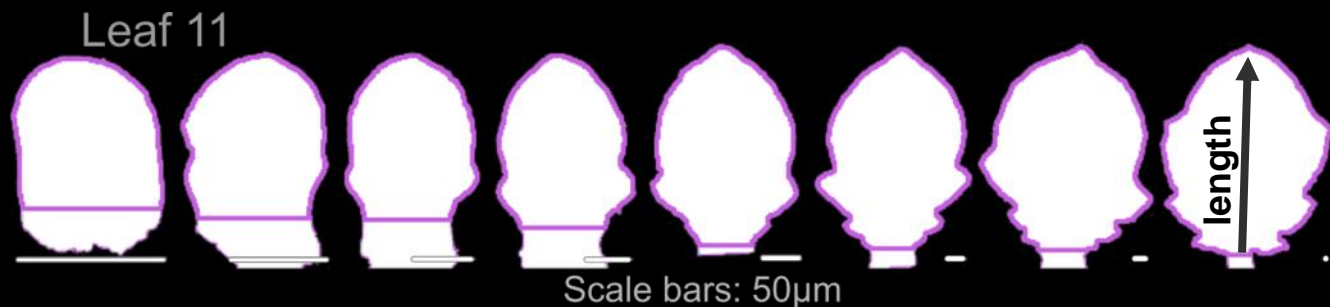
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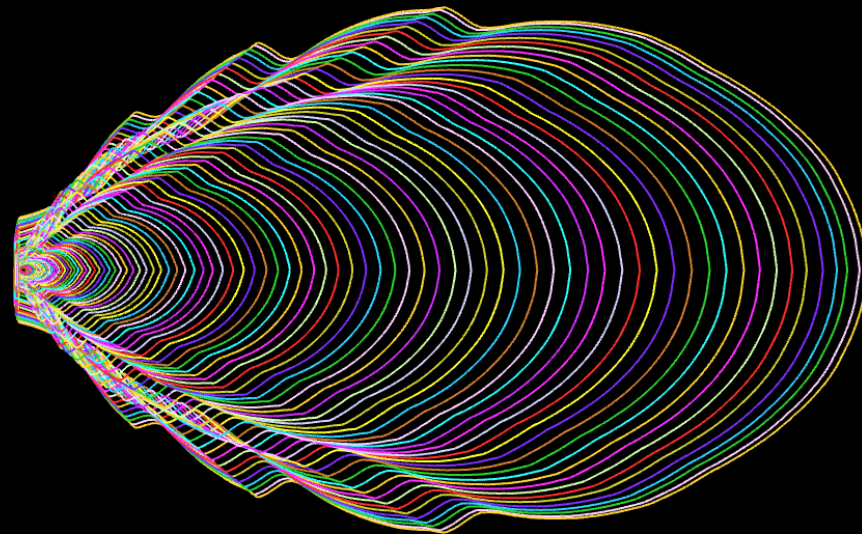
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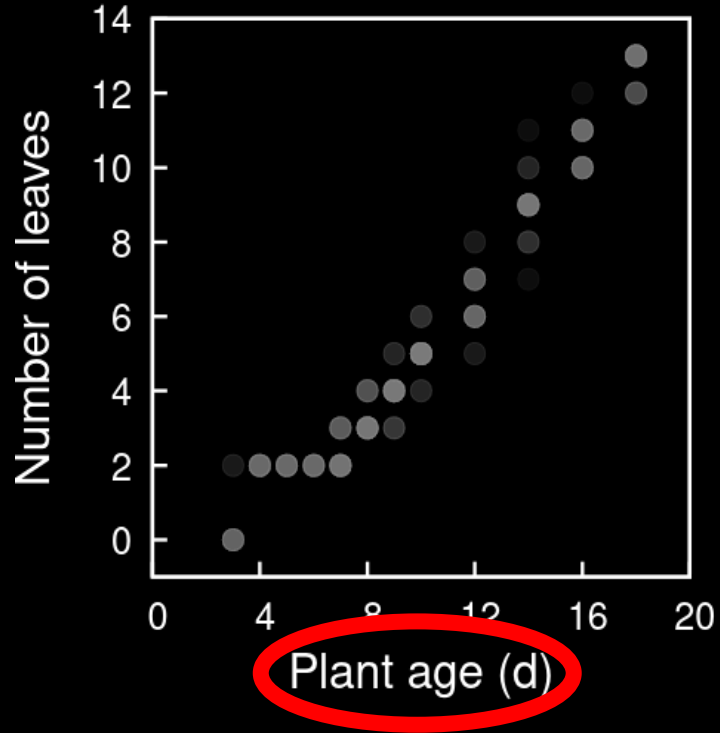


This is not the real  
temporal dynamics!  
Time should be  
reintroduced  
 $\Rightarrow$  “dating” leaves



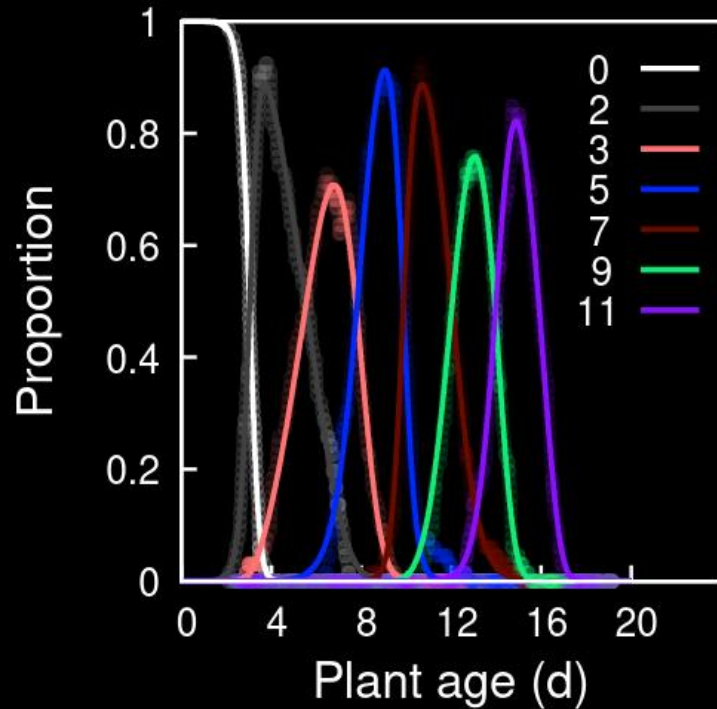
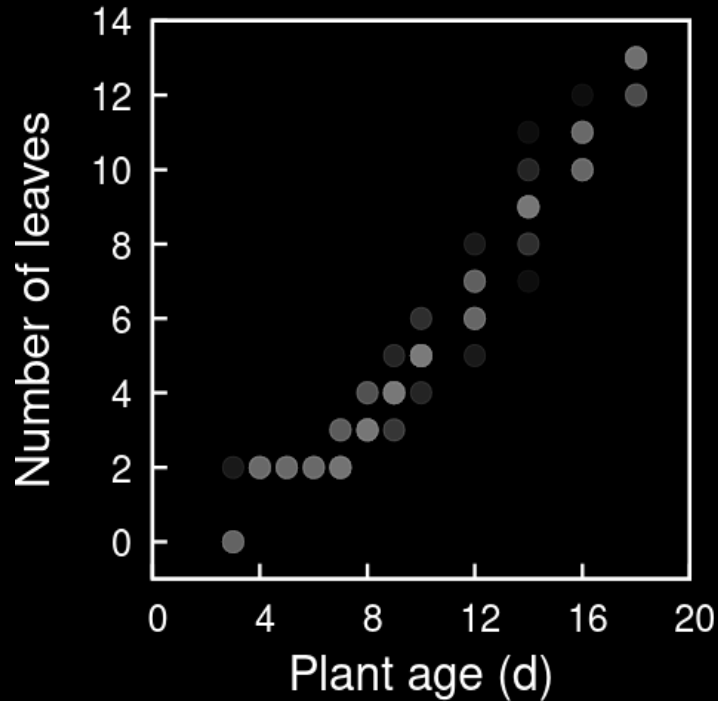


## First : determining the starting time using modeling



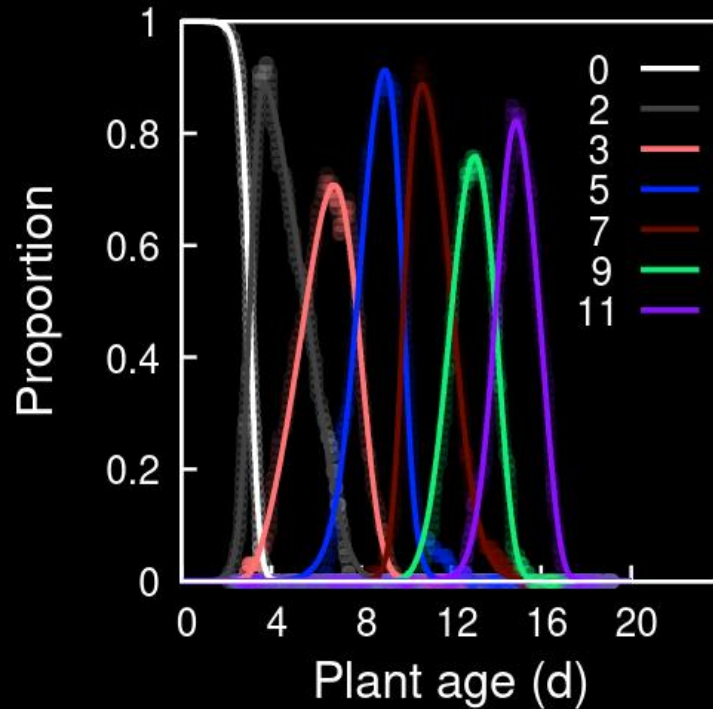
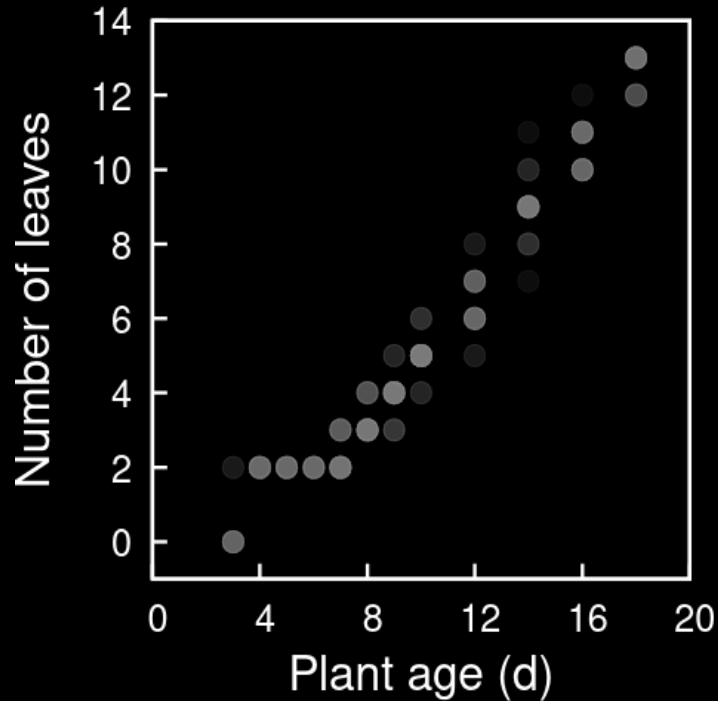
Available time: plant age

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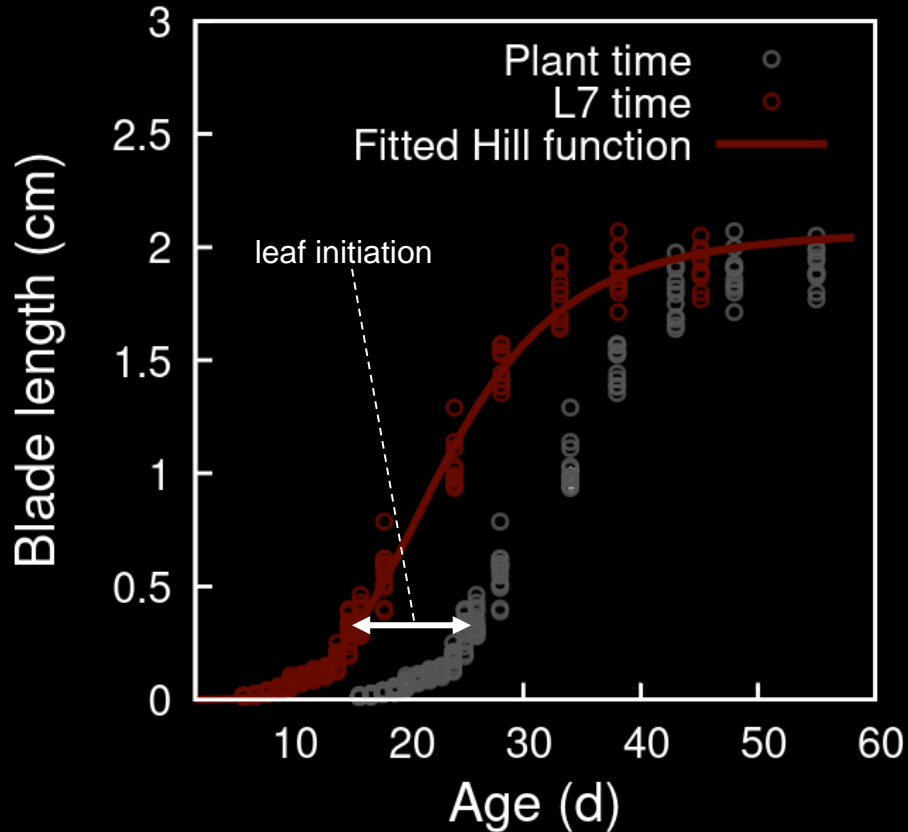
$$\frac{dN_k(t)}{dt} = \alpha_{k-1}(t)N_{k-1}(t) - \alpha_k(t)N_k(t)$$

## First : determining the starting time using modeling



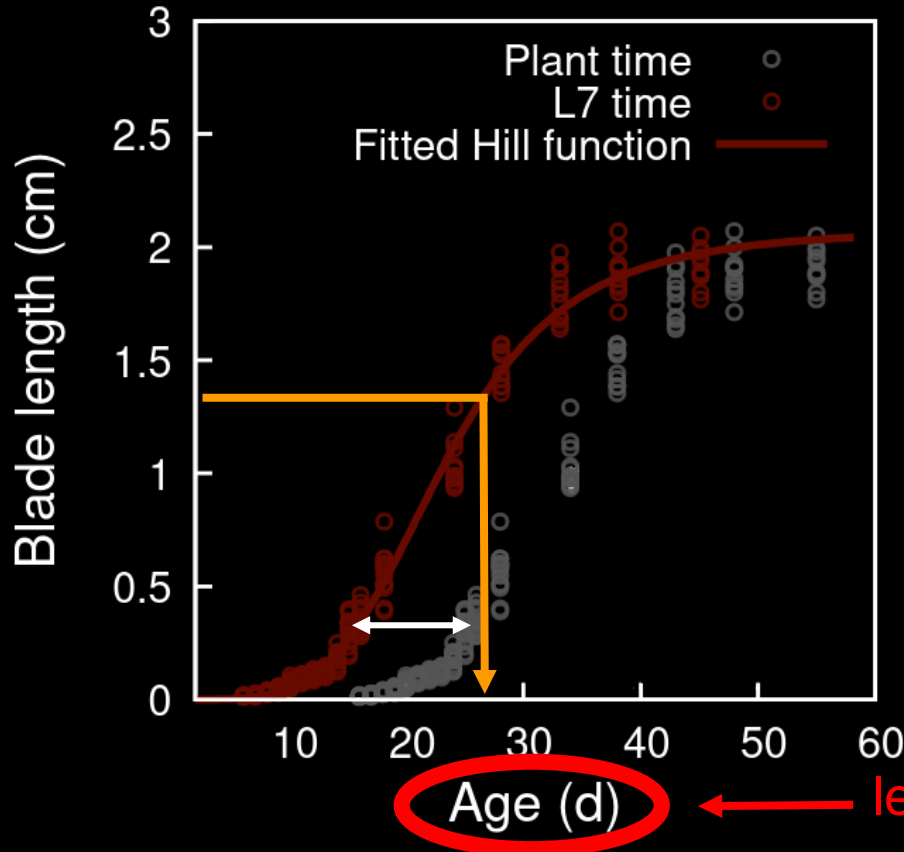
Modeling strategy useable to estimate initiation time of any structures appearing sequentially

## Second: relying organ size and age



- Leaf size is recorded as a function of plant age
- Time translation is applied to get measures as a function of leaf age
- A sigmoid (Hill function) is fitted to the data

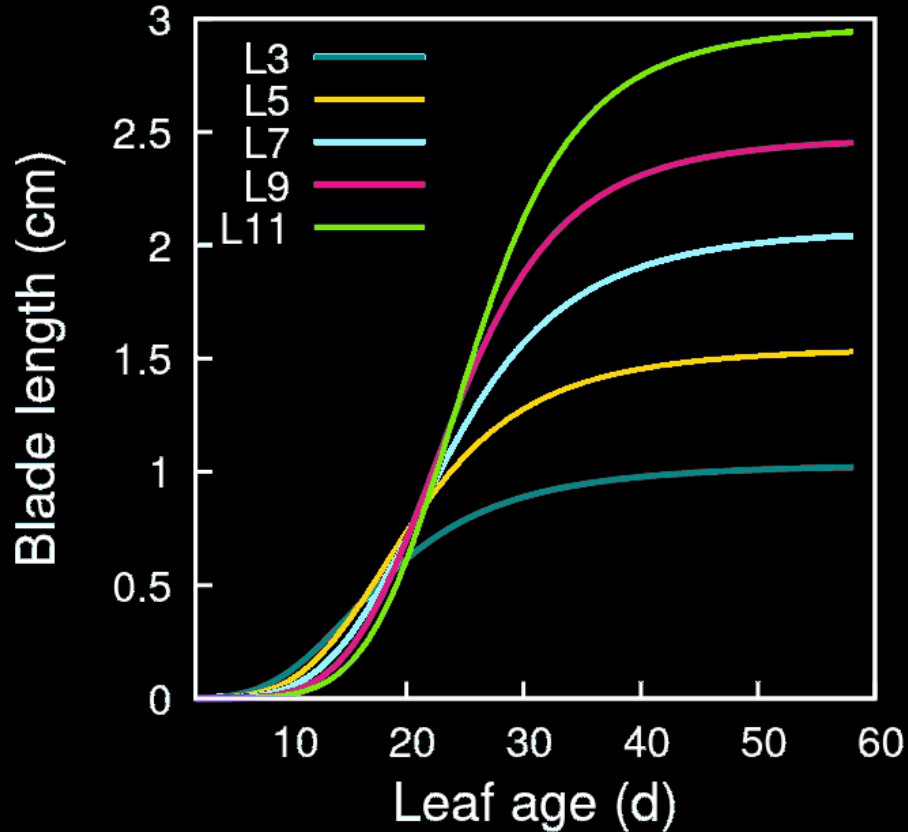
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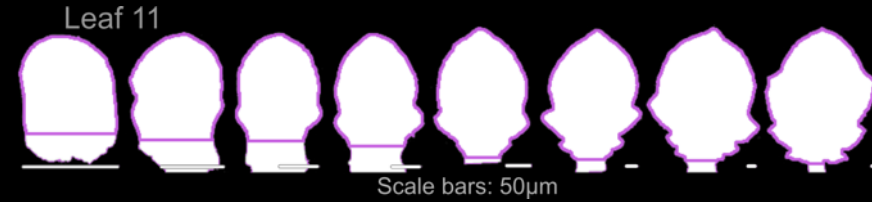
The organ age can be retrieved from its size

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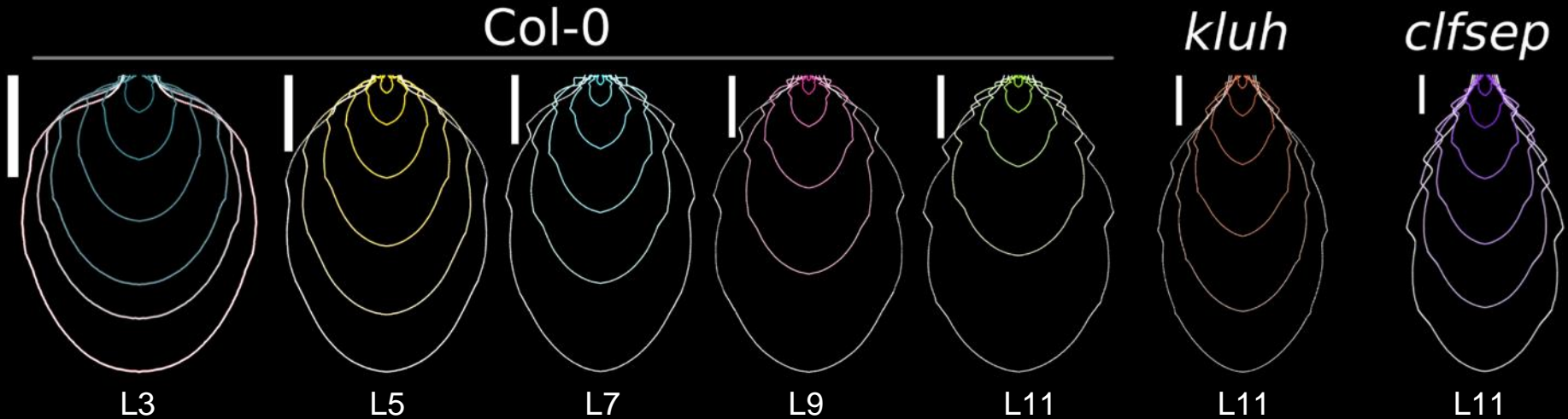
# Reconstructing temporal growth trajectories



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The continuous organ shape evolution is retrieved from fixed images only

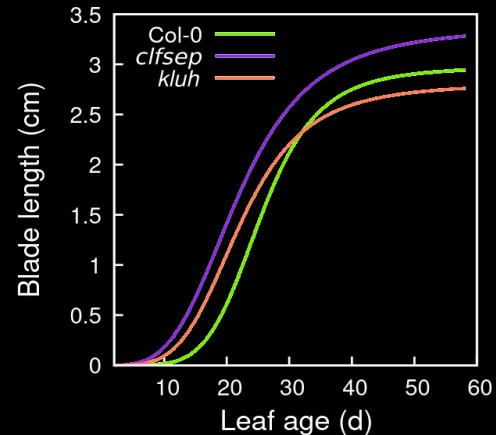
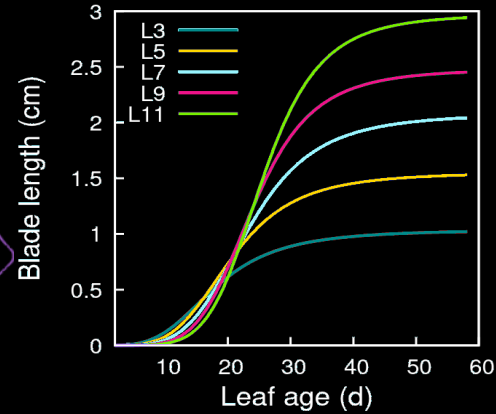
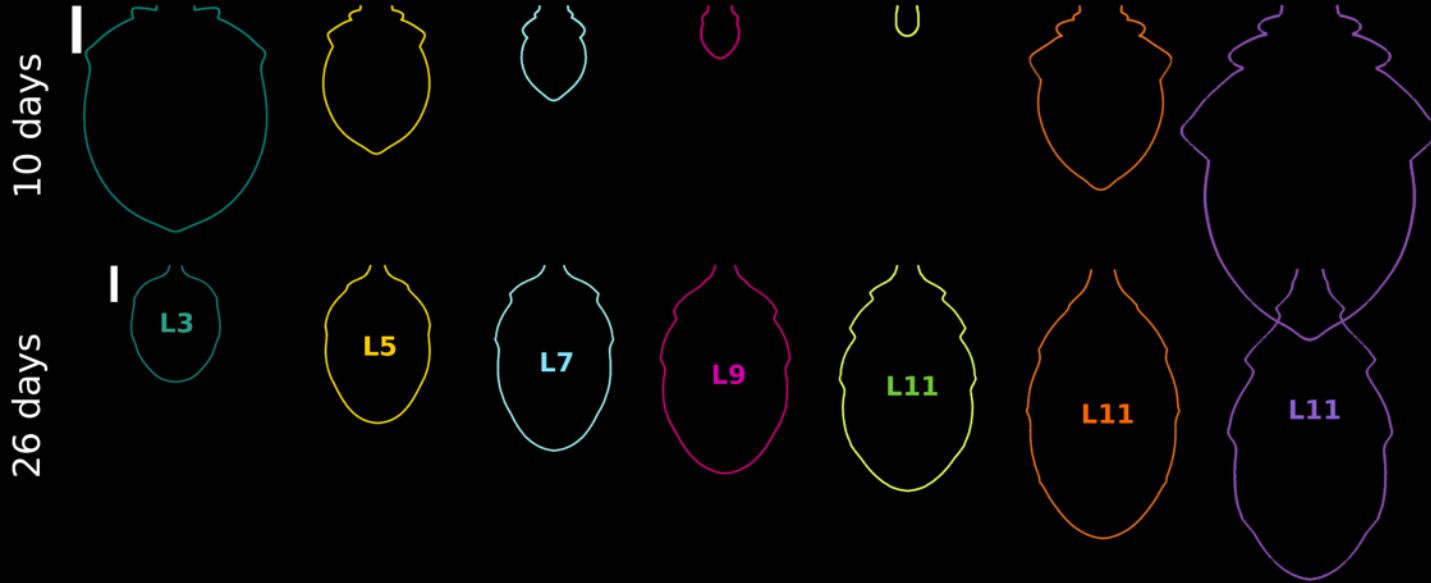
# Reconstructing temporal growth trajectories



Contours of leaves at 220h, 290h, 360h, 430h, 500h and 570h  
after initiation



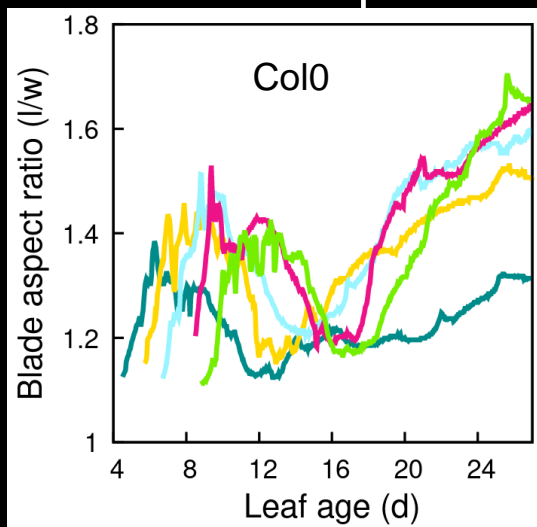
# Reconstructing temporal growth trajectories



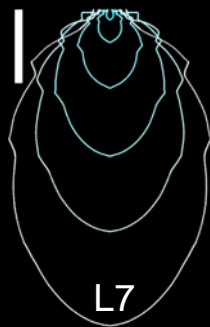
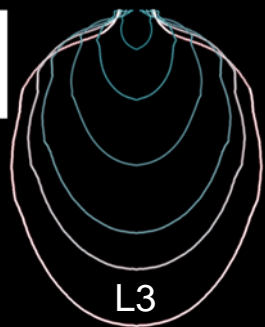
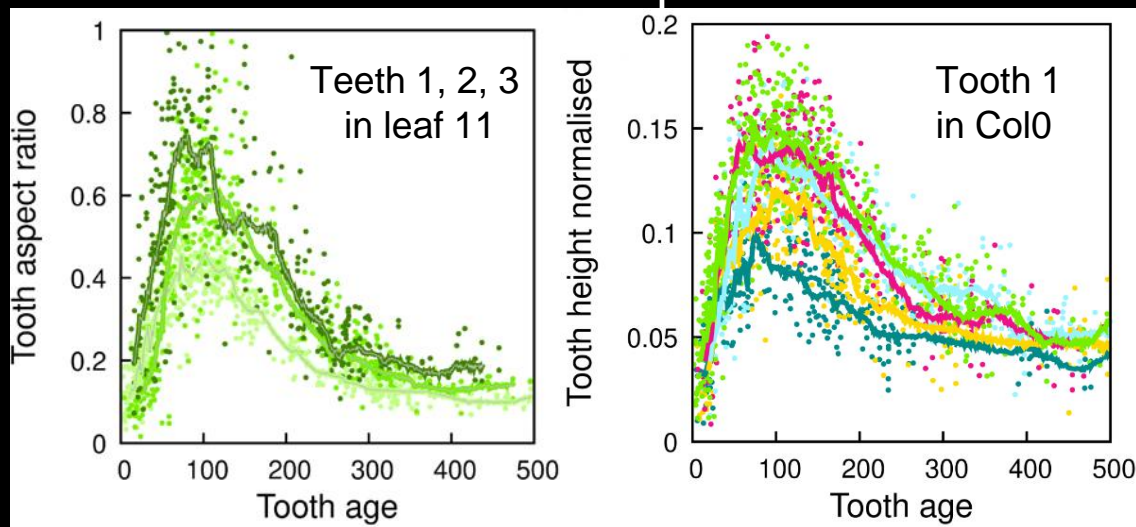
Whole growth has to be considered to fully understand developmental kinetics

# Quantifying shapes during growth

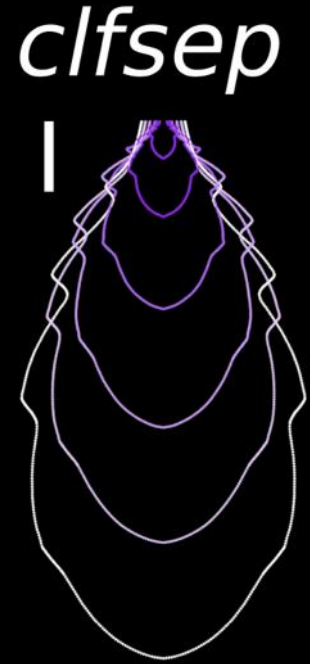
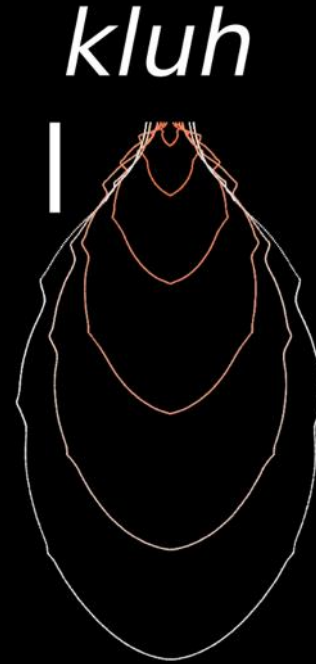
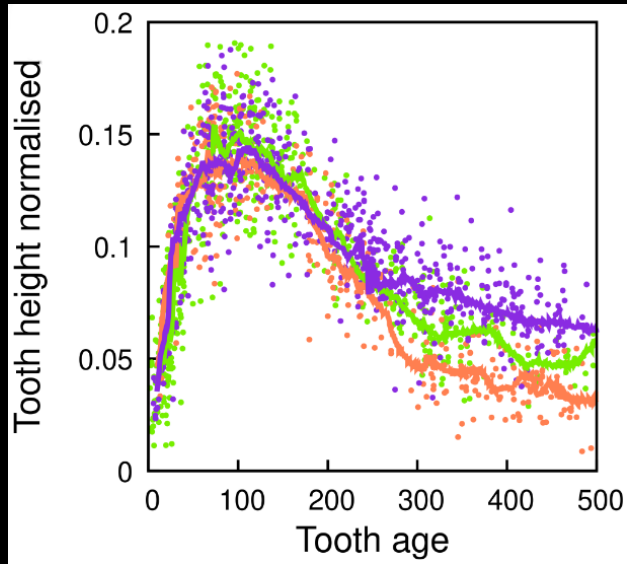
Global shape



Local shape



# Revealing key events during the development



## Static data used to reconstruct morphodynamics

- a model to determine apparition times (simple counting protocols)
- a fitted growth function relating size and age
- explicit long-term morphodynamics can be retrieved/quantified
- the effects of alterations (mutation, environment, ...) can be localized/quantified during all growth

In *Arabidopsis thaliana*:

- leaf heteroblasty was characterized during growth (global/local scales)
- dynamics of successive leaves in *Arabidopsis thaliana* follow a same program, with graded parameters
- key developmental events can be revealed in space and time (using, e.g., mutants)

# A collaborative work at IJPB, INRAE Versailles

## Team Modeling and Digital Imaging

- Philippe ANDREY
- Eric BIOT
- Mohamed OUGHOU

## Team Transcription Factors and Architecture

- Nicolas ARNAUD
- Patrick LAUFS
- Aude MAUGARNY-CALES